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Authors: Provides an updated overview on efficacy of biocidal active substances and antiseptics Explores the potential of those agents to enhance tolerance and maintain the efficacy of useful antiseptics 8131 Accesses 3 Citations This updated and expanded
second edition of Antiseptic Stewardship serves as a comprehensive reference guide to common biocidal active substances and antiseptic agents, as well as cross-resistance to antibiotics. In addition, the book discusses the
appropriate and targeted use of biocidal active substances by balancing their expected health benefits against the likelihood of clinically relevant resistance, including misuse and overuse of some products, helps readers make informed decisions
about disinfectants and antiseptic products based on their composition. Various biocidal active substances and antiseptic agents are used for disinfection and household products. However, not all of them provide significant health benefits, especially for some products used in
human medicine. Antimicrobial soaps, surface disinfectants, instrument disinfectants and wound antiseptics may contain one or more biocidal active ingredients with comparable antimicrobial efficacy, but with large differences in their potential for microbial adaptation and tolerance. Increased bacterial tolerance has been described for several
biocidal active substances and antiseptics, sometimes including cross-resistance to antibiotics. The book is therefore intended to help reduce unnecessary selection pressure on emerging pathogens, including by describing non-biocidal alternatives for specific antimicrobial applications, with the aim of retaining the powerful biocidal agents and
antiseptics for those applications where there is a clear health benefit (e.g. reduction of healthcare, industrial and veterinary professionals as well as educated laypersons interested in efficient and controlled disinfection strategies. Günter Kampf is an Associated Professor for Hygiene and
Environmental Medicine at the University of Greifswald, Germany. He has published more than 250 mostly international articles on various aspects of infection control, such as hand hygiene, surface disinfection and the epidemiology of COVID-19. He has published so far 11 textbooks on hand hygiene, surface hygiene and the epidemiology of COVID-19.
19. As a recognized medical specialist in hygiene and environmental medicine, he worked for a manufacturer of chemical disinfectants for 18 years, the last 5 years of which as Director of Science. Since 2016 he has been self-employed and continues his scientific work together with infection control consultancy for hospitals, medical practices and
for this Substance1-Decanaminium, N-decyl-N,N-dimethyl-, chloride (1:1)Ammonium, N-decyl-N,N-dimethyl-, chlorideDidecyldimethyl-, chlorideDidecyldim
1094740-76-7, 1632379-58-8 Cosmetics and personal care products are not required to be tested for safety before being allowed on the market. The Skin Deep® scoring system was designed to help the public understand whether a product is safe to use or whether it contains ingredients of concern. Every product and ingredient in Skin Deep gets a
two-part score - one for hazard and one for data availability. The safest products score well by both measures, with a low hazard rating and a fair or better data availability rating. HOW WE DETERMINE SCORES The Skin Deep ingredient hazard score, from 1 to 10, reflects known and suspected hazards linked to the ingredients. The EWG Verified®
mark means a product meets EWG's strictest criteria for transparency and health. The Skin Deep data availability rating reflects the number of scientific literature. NONE LIMITED FAIR GOOD ROBUST Didecyldimethylammonium chloride Names Preferred IUPAC name N-Decyl-N,N-
dimethyldecan-1-aminium chloride[1] [2] 1-Decanaminium[1] N-decyl-N,N-dimethyl-, chloride[1] Didecyldimethyl-, chloride[1] Did
UHFFFAOYSA-M YInChI=1/C22H48N.ClH/c1-5-7-9-11-13-15-17-19-21-23(3,4)22-20-18-16-14-12-10-8-6-2;/h5-22H2,1-4H3;1H/q+1;/p-1Key: RUPBZQFQVRMKDG-REWHXWOFAJ SMILES [Cl-].C(CCCCC(N+)(CCCCCCCCN)C)CCCC Properties Chemical formula C22H48ClN Molar mass 362.08 g/mol Appearance liquid[3] Density 0.87 g/cm3 (20 °C)
[3] Pharmacology ATC code D08A[06 (WHO) Hazards Occupational safety and health (OHS/OSH): Main hazards corrosive[3] GHS labelling: Pictograms Except where otherwise noted, data are given for materials in their standard state (at 25 °C [77 °F], 100 kPa). Y verify (what is YN?) Infobox references Chemical compound
Didecyldimethylammonium chloride (DDAC) is a quaternary ammonium compound used as an antiseptic/disinfectant. It causes the disruption of intermolecular interactions and the dissociation of lipid bilayers. The bacteriostatic (prevent growth) or bacteriostatic (
of the microbial population.[4] It is a broad spectrum biocidal against bacteria and fungi and can be used as disinfectant cleaner for linen, recommended for use in hospitals, hotels and industries. It is also used in gynaecology, surgery, ophthalmology, pediatrics, OT, and for the sterilization of surgical instruments, endoscopes and surface disinfection.
In mice this disinfectant was found to cause infertility and birth defects when combined with alkyl (60% C14, 25% C16) dimethyl benzyl ammonia compounds which was reviewed by the U.S. Environmental Protection Agency (U.S. EPA)
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use the material. Please enable Javascript in order to use PubChem website. Stereochemistry ACHIRAL Molecular Formula C22H48N.Cl Molecular Weight 362.076 Optical Activity NONE Defined Stereochemistry ACHIRAL Molecular Formula C22H48N.Cl Molecular Weight 362.076 Optical Activity NONE Defined Stereochemistry ACHIRAL Molecular Formula C22H48N.Cl Molecular Formula C
UHFFFAOYSA-M InChI=1S/C22H48N.ClH/c1-5-7-9-11-13-15-17-19-21-23(3,4)22-20-18-16-14-12-10-8-6-2;/h5-22H2,1-4H3;1H/q+1;/p-1 HIDE SMILES / InChI DIDECYLDIMONIUM (as a salt, didecyldimethylammonium chloride (DDAC)) is used in many types of biocidal products including tableware, carpets, humidifiers, and swimming pools. It shows a
broad spectrum of activity against both gram-positive and gram-negative bacteria and is also effective on fungi and viruses, including those that are enveloped. Name English Common Name 
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NLM provides access to scientific literature. Inclusion in an NLM database does not imply endorsement of, or agreement with, the contents by NLM or the National Institutes of Health. Learn more: PMC Disclaimer | PMC Copyright Notice . Author manuscript; available in PMC: 2016 Jul 26. Didecyldimethylammonium chloride (DDAC) is a dialkyl-
quaternary ammonium compound that is used in numerous products for its bactericidal, virucidal and fungicidal properties. There have been clinical reports of immediate and delayed hypersensitivity reactions in exposed individuals; however, the sensitization potential of DDAC has not been thoroughly investigated. The purpose of these studies was
to evaluate the irritancy and sensitization potential of DDAC following dermal exposure in a murine model. DDAC induced significant irritancy (0.5 and 1%), evaluated by ear swelling in female Balb/c mice. Initial evaluation of the sensitization potential was conducted using the local lymph node assay (LLNA) at concentrations ranging from 0.0625-1%
A concentration-dependent increase in lymphocyte proliferation was observed with a calculated EC3 value of 0.17%. Dermal exposure to DDAC did not induce increased production of IgE as evaluated by phenotypic analysis of draining lymph node B-cells (IgE+B220+) and measurement of total serum IgE levels. Additional phenotypic analysis
revealed significant and dose-responsive increases in the absolute number of B-cells, CD4+ T-cells and dendritic cells in the draining lymph nodes, along with significant increases in the percentage of B-cells (0.25% and 1% DDAC) at Day 10 following 4 days of dermal exposure. There was also a significant and dose-responsive increases
in the number of activated CD44 + CD4 + and CD8+ T-cells and CD8+ B-cells and CD86+ B-cells and dendritic cells following exposure to all concentrations of DDAC. These results demonstrate the potential for development of irritation and hypersensitivity responses to DDAC following dermal exposure and raise concerns about the use of this chemical and other
quaternary ammonium compounds that may elicit similar effects. Keywords: DDAC, hypersensitivity, allergic disease, quaternary ammonium compounds (QAC) are commonly used as water-based surface disinfectants due to their low volatility and they are increasingly being used in hospitals, hotels and in consumer
products (Zhang et al. 2015) due to their broad antimicrobial capabilities. In healthcare settings for the decontamination of surgical instruments, endoscopes and other medical instruments, endoscopes and other medical instruments, endoscopes and other medical instruments.
consumer products that are utilized orally (mouthwash) or applied to the skin or eyes for the purpose of decreasing microbial contamination and reducing the incidence of pathogen-induced illness. All QAC are permanently charged ions with four alkyl side chains. Their structures contain at least one hydrophobic hydrocarbon chain linked to a
positively charged nitrogen atom and other alkyl groups that are mostly short-chain substitutes such as methyl or benzyl groups. The biocidal activity is conferred through alkyl chain length (McBain et al. 2004). The ability to adapt and optimize QAC structure to target specific microbial species has recently increased the utilization of these
compounds for use in consumer products (Carson et al. 2008). Dialkyl QAC represent the newest generation of QAC and exhibit a wide spectrum of activity. These new synthetic polymeric QAC contain multiple positively-charged amine centers that confer anti-microbial, antistatic, and surfactant properties in solution. One of the newer QAC in
common use is didecyldimethylammonium chloride (DDAC). DDAC is a broad-spectrum bactericidal and fungicidal biocide that exhibits antimicrobial activity against several pathogens such as Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa (Walsh et al. 2003), Legionella pneumophilia (Skaliy et al. 1980), Stachybotrys chartarum
and enveloped and non-enveloped viruses (Argy et al. 1999). It is used in several types of applications including: industrial processes, swimming pools and aquatic areas, wood treatment, healthcare and food handling and storage (Ohnuma et al. 2011). QAC have been used for over 50 years and have generally been regarded as safe; however, there is
very limited published research describing the toxicity of these compounds, especially regarding the newer formulations such as DDAC. A study conducted by Ohnuma et al. (2011) examined the pulmonary defense system following a single intratracheal instillation of DDAC (60 and 150 µg/kg) in C57BL/6J mice. Those authors found that exposure to
the high dose induced lung injury as early as 1-d post-exposure, as evidenced by increased lactate dehydrogenase (LDH) activity and protein concentrations in the BAL (specifically macrophages, neutrophils and lymphocytes), along with increases in interleukin (IL)-6
production by 7-days post-exposure. The authors also suggested that DDAC exposure with lipopolysaccharide (LPS) generated a further enhancement in pulmonary inflammation suggesting a potential increase in susceptibility to
bacterial agents. A more recent study investigated the effects of DDAC following a 2-week inhalation exposure (Lim & Chung 2014) in Sprague-Dawley rats. Aside from decreases in body weight at the highest exposure concentration (3.6 mg/m3), no changes in hematological and blood biochemistry parameters were observed and mild changes were
observed in BAL cell counts and cell damage parameters. Interestingly, a recent study observed that, following the introduction of DDAC in an animal facility there was decreased reproductive performance of the laboratory mice. Additional examination identified decreases in fertility and fecundity including increased time to first litter, longer
pregnancy intervals, fewer pups per litter and fewer pregnancies in mice following DDAC exposure (Melin et al. 2014). Epidemiological data and case studies indicate that healthcare workers have an elevated risk for development of sensitization and allergic asthma from either dermal or inhalation exposure to chemicals compared to non-healthcare workers have an elevated risk for development of sensitization and allergic asthma from either dermal or inhalation exposure to chemicals compared to non-healthcare workers have an elevated risk for development of sensitization and allergic asthma from either dermal or inhalation exposure (Melin et al. 2014).
workers (Warshaw et al. 2008). Biocides such as QAC have been identified to be among the most common allergens in the healthcare profession (Bernstein et al. 2014). A study evaluating 142 patients with suspected allergies to the most commonly used QAC, i.e
benzalkonium chloride (BAC) and benzethonium chloride (BEC), confirmed sensitization by patch test to these compounds in 20% of the subjects who tested positive (Dao et al. 2012). While there have been fewer overall reported cases of sensitization to the newer
formulations of QAC, allergic contact dermatitis and immediate-type allergic reactions caused by DDAC exposure have been recently reported. Four cases, which were confirmed by patch testing or open epicutaneous tests, describe contact dermatitis presenting in hospital and laboratory workers on the hands/wrists and face following exposure to
DDAC present in a disinfectant (Dejobert et al. 1997; Dibo & Brasch 2001; Ruiz Oropeza et al. 2011). In addition, there is also a confirmed case of immediate allergy reported in an
individual in catering school who was exposed to DDAC in a cleaning product. This individual suffered from urticaria, facial angioedema and dyspnea within 10 min of exposure. A 20-min open epicutaneous test with the diluted cleaning product and DDAC confirmed immediate-type hypersensitivity (Houtappel et al. 2008). All symptoms in the
individuals resolved when exposure to DDAC was eliminated. While these studies suggest a role for QACs and DDAC in allergic disease, the exact mechanism of action for sensitization to these compounds remains to be investigated and explained. Due to the high potential for human exposure, epidemiological studies suggesting an association with
allergic disease and the lack of dermal toxicological data, this study was performed to evaluate the irritancy and skin sensitization potential of DDAC using a murine model in an effort to assess its role in the development of allergic disease. Female BALB/c mice were used for the murine models. This mouse strain has a T-helper (TH)-2 bias and is
commonly used to evaluate IgE-mediated sensitization (Woolhiser et al. 2000; Klink & Meade 2003). The mice were purchased from Taconic (Germantown, NY) at 6-8 weeks-of-age. Upon arrival, the animals were allowed to acclimate for a minimum of 5 days. Each shipment of animals was randomly assigned to a treatment group, weighed and
individually identified (via tail marking) using a permanent marker or tattoo. A preliminary analysis of variance on body weights was performed to ensure a homogeneous distribution of animals were housed at a maximum of 5/cage in ventilated plastic shoebox cages with hardwood chip bedding. NIH-31 modified
6% irradiated rodent diet (Harlan Teklad) and tap water were provided from water bottles, ad libitum. The animal facility temperature was maintained on 12-h intervals. All animal experiments were performed in the Association for Assessment and Accreditation
of Laboratory Animal Care (AAALAC) accredited NIOSH animal facility in accordance with an animal protocol approved by the Institutional Animal Care (CAS#67-64-1), α-Hexylcinnamaldehyde (HCA; CAS#101-86-0) and 2,4-toluene diisocyanate
(TDI; CAS#584-84-9) were purchased from Sigma-Aldrich (Milwaukee, WI). Chemical structure of DDAC to be used for dermal exposures. Mice (n = 3/group) were exposed topically to acetone vehicle or increasing concentrations of DDAC (up to 40%) in
acetone on the dorsal surface of each ear (25 µl per ear) for three consecutive days. Acetone was selected as the appropriate vehicle based on solubility, historical control data and accepted use in skin sensitization studies (NIEHS 1999). Animals were allowed to rest for 2 days following the final exposure and then weighed and examined for signs of
overt toxicity including loss of body weight, fatique/lack of activity and ungroomed fur. To determine the irritancy and sensitization potential of DDAC, a combined LLNA was conducted as previously described in the ICCVAM Peer Review Panel report (NIEHS 1999) with minor
modifications. Briefly, mice (n = 5/group) were exposed topically to acetone vehicle, increasing concentrations of test agent or positive control (HCA) on the dorsal surface of each ear (25 µl per ear) for three consecutive days. HCA (30%) is an accepted and well-characterized positive control for the LLNA (NIEHS 1999). For irritancy evaluation, the
thickness of the right and left ear pinnae of each mouse was measured using a modified engineer micrometer (Mitutoyo Co., Tokyo, Japan) before the first chemical administration and 24 h following the final exposure. The mean percentage of ear swelling was calculated based on the following equation: [(mean post-challenge ear thickness — mean
pre-challenge ear thick-ness)/mean pre-challenge thickness] × 100. Animals were allowed to rest for 2 days following the final exposure. On Day 6, mice were injected intravenously via the lateral tail vein with 20 μCi [3H]-thymidine (Dupont NEN; specific activity = 2 Ci/mmol). Five hours after [3H]-thymidine injection, the animals were euthanized via
CO2 inhalation and the left and right auricular draining lymph nodes (DLN; drain site of chemical application) located at the bifurcation of the jugular vein were excised and pooled for each animal. Single cell suspensions were made and incubated overnight in 5% trichloroacetic acid and samples were then counted in a Packard Tri-Carb 2500TR
liquid scintillation analyzer (Perkin Elmer, Waltham, MA). Stimulation indices (SI) were calculated by dividing the mean disintegrations per minute (DPM) per test group by the mean DPM for the vehicle control group. EC3 values (concentration of chemical required to induce a 3-fold increase over the vehicle control) were calculated based on the
equation from Basketter et al. (1999). Dosing concentration of chemical required to induce a 3-fold increase over the vehicle control (EC3) was calculated based on the equations from Basketter et al. (1999). For the phenotypic
analysis, mice (n = 5/group) were exposed to 25 ml/ear of the acetone vehicle, increasing concentrations of test article (0.25%, 0.5% and 1% DDAC) or positive control (1.0% TDI) once daily for four consecutive days. Animals were euthanized by CO2 inhalation on Day 10, weighed and examined for gross pathology. The liver, spleen, kidneys and
thymus were removed, cleaned of connective tissue and weighed. Serum was collected for total IgE analysis (see below). DLN cell suspensions (2 nodes/animal/3 ml PBS) were prepared by mechanical disruption of tissues between frosted microscope slides in phosphate buffered saline (PBS) and counted on a Cellometer (Nexcelom Bioscience,
Lawrence, MA). Cells (1-2 × 106) were aliquoted into a 96-well U-bottom plates and washed in staining buffer containing anti-mouse CD16/32 antibody (clone 2.4G2) for blocking of Fc receptors (BD Biosciences, San Jose, CA). Cells were next incubated
with a cocktail of flurochrome-conjugated antibodies specific for cell surface antigens, including IgE-FITC (R35-72), B220-V500 (RA3-6B2), CD86-APC (GL1), MHC II-Alexa Fluor F700 (M5/11.15.2) and CD44-eFluor 780 (IM7) (eBioscience, San
Diego, CA), Cells were then washed in staining buffer and then fixed in Cytofix buffer according to manufacturer instructions (BD Biosciences). Within 24 h, the cells were re-suspended in staining buffer and 50 000 events collected and analyzed on an LSR II flow cytometer (BD Biosciences). Compensation controls were prepared with OneComp
eBeads (eBioscience). The IgE+B220+ populations were analyzed as described by Manetz and Meade (1999). Data analysis was performed using FlowJo 7.6.5 software (TreeStar Inc., Ashland, OR). Following euthanasia of animals included in the phenotyping study, blood samples were collected via cardiac puncture. Sera were separated by
centrifugation and frozen at -20°C for subsequent analysis of total IgE by ELISA. A standard colorimetric sandwich ELISA was performed as previously described (Anderson et al. 2007). For analysis of variance (ANOVA) was conducted. If
the ANOVA showed significance at p
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